PATENT COOPERATION TREATY

From the INTERNATIONAL BUREAU

PCT

NOTIFICATION OF ELECTION

(PCT Rule 61.2)

To:

Assistant Commissioner for Patents
United States Patent and Trademark Office
Box PCT
Washington, D.C.20231
ETATS-UNIS D'AMERIQUE

in its capacity as elected Office

Date of mailing (day/month/year) 25 October 2000 (25.10.00)	
International application No. PCT/US00/06829	**Applicant's or agent's file reference** 440202/PALL
International filing date (day/month/year) 15 March 2000 (15.03.00)	**Priority date** (day/month/year) 16 March 1999 (16.03.99)
Applicant BORMANN, Thomas, J. et al	

1. The designated Office is hereby notified of its election made:

[X] in the demand filed with the International Preliminary Examining Authority on:

28 September 2000 (28.09.00)

[ ] in a notice effecting later election filed with the International Bureau on:

2. The election [X] was

[ ] was not

made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO **34, chemin des Colombettes** **1211 Geneva 20, Switzerland**	Authorized officer Claudio Borton
Facsimile No.: (41-22) 740.14.35	Telephone No.: (41-22) 338.83.38

PATENT COOPERATION TREATY

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Date of mailing (day/month/year) 25 October 2000 (25.10.00)	
International application No. PCT/US00/06829	**Applicant's or agent's file reference** 440202/PALL
International filing date (day/month/year) 15 March 2000 (15.03.00)	**Priority date** (day/month/year) 16 March 1999 (16.03.99)
Applicant BORMANN, Thomas, J. et al	

1. The designated Office is hereby notified of its election made:

[X] in the demand filed with the International Preliminary Examining Authority on:

28 September 2000 (28.09.00)

[ ] in a notice effecting later election filed with the International Bureau on:

2. The election [X] was

[ ] was not

made before the expiration of 19 months from the priority date or, where Rule 32 applies, within the time limit under Rule 32.2(b).

The International Bureau of WIPO
34, chemin des Colombettes
1211 Geneva 20, Switzerland

Facsimile No.: (41-22) 740.14.35

Authorized officer

Claudio Borton

Telephone No.: (41-22) 338.83.38

PATENT COOPERATION TREATY

From the
INTERNATIONAL PRELIMINARY EXAMINING AUTHORITY

To:

JAY, U.
LEYDIG, VOIT & MAYER
700 Thirteenth Street, N.W.
Suite 300
Washington, D.C. 20005
ETATS-UNIS D'AMERIQUE

PCT

NOTIFICATION OF TRANSMITTAL OF THE INTERNATIONAL PRELIMINARY EXAMINATION REPORT
(PCT Rule 71.1)

Date of mailing *(day/month/year)*	29.06.2001
Applicant's or agent's file reference 440202/PALL	**IMPORTANT NOTIFICATION**

International application No.	International filing date *(day/month/year)*	Priority date *(day/month/year)*
PCT/US00/06829	15/03/2000	16/03/1999

Applicant
PALL CORPORATION

1. The applicant is hereby notified that this International Preliminary Examining Authority transmits herewith the international preliminary examination report and its annexes, if any, established on the international application.

2. A copy of the report and its annexes, if any, is being transmitted to the International Bureau for communication to all the elected Offices.

3. Where required by any of the elected Offices, the International Bureau will prepare an English translation of the report (but not of any annexes) and will transmit such translation to those Offices.

4. **REMINDER**

The applicant must enter the national phase before each elected Office by performing certain acts (filing translations and paying national fees) within 30 months from the priority date (or later in some Offices) (Article 39(1)) (see also the reminder sent by the International Bureau with Form PCT/IB/301).

Where a translation of the international application must be furnished to an elected Office, that translation must contain a translation of any annexes to the international preliminary examination report. It is the applicant's responsibility to prepare and furnish such translation directly to each elected Office concerned.

For further details on the applicable time limits and requirements of the elected Offices, see Volume II of the PCT Applicant's Guide.

Name and mailing address of the IPEA/	Authorized officer
European Patent Office D-80298 Munich Tel. +49 89 2399 - 0 Tx: 523656 epmu d Fax: +49 89 2399 - 4465	Ipinazar, P Tel.+49 89 2399-8131

Form PCT/IPEA/416 (July 1992)

09/936732

PATENT COOPERATION TREATY

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REC'D 03 JUL 2001
WIPO PCT

INTERNATIONAL PRELIMINARY EXAMINATION REPORT

(PCT Article 36 and Rule 70)

14

Applicant's or agent's file reference	FOR FURTHER ACTION	See Notification of Transmittal of International Preliminary Examination Report (Form PCT/IPEA/416)
440202/PALL		
International application No.	International filing date *(day/month/year)*	Priority date *(day/month/year)*
PCT/US00/06829	15/03/2000	16/03/1999
International Patent Classification (IPC) or national classification and IPC B01D39/16		
Applicant PALL CORPORATION		

1. This international preliminary examination report has been prepared by this International Preliminary Examining Authority and is transmitted to the applicant according to Article 36.

2. This REPORT consists of a total of 5 sheets, including this cover sheet.

   ☒ This report is also accompanied by ANNEXES, i.e. sheets of the description, claims and/or drawings which have been amended and are the basis for this report and/or sheets containing rectifications made before this Authority (see Rule 70.16 and Section 607 of the Administrative Instructions under the PCT).

   These annexes consist of a total of 4 sheets.

3. This report contains indications relating to the following items:

   - I ☒ Basis of the report
   - II ☐ Priority
   - III ☐ Non-establishment of opinion with regard to novelty, inventive step and industrial applicability
   - IV ☐ Lack of unity of invention
   - V ☒ Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations suporting such statement
   - VI ☐ Certain documents cited
   - VII ☒ Certain defects in the international application
   - VIII ☒ Certain observations on the international application

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Date of submission of the demand	Date of completion of this repor
28/09/2000	29.06.2001
Name and mailing address of the international preliminary examining authority: European Patent Office D-80298 Munich Tel. +49 89 2399 - 0 Tx: 523656 epmu d Fax: +49 89 2399 - 4465	Authorized officer Polesak, H Telephone No. +49 89 2399 8628

Form PCT/IPEA/409 (cover sheet) (January 1994)

I. Basis of th report

1. With regard to the **elements** of the international application *(Replacement sheets which have been furnished to the receiving Office in response to an invitation under Article 14 are referred to in this report as "originally filed" and are not annexed to this report since they do not contain amendments (Rules 70.16 and 70.17))*:
Description, pages:

1-26	as originally filed	

Claims, No.:

1-30	with telefax of	26/02/2001

Drawings, sheets:

1/5-5/5	as originally filed

2. With regard to the **language**, all the elements marked above were available or furnished to this Authority in the language in which the international application was filed, unless otherwise indicated under this item.

   These elements were available or furnished to this Authority in the following language: , which is:

   - ☐ the language of a translation furnished for the purposes of the international search (under Rule 23.1(b)).
   - ☐ the language of publication of the international application (under Rule 48.3(b)).
   - ☐ the language of a translation furnished for the purposes of international preliminary examination (under Rule 55.2 and/or 55.3).

3. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international preliminary examination was carried out on the basis of the sequence listing:

   - ☐ contained in the international application in written form.
   - ☐ filed together with the international application in computer readable form.
   - ☐ furnished subsequently to this Authority in written form.
   - ☐ furnished subsequently to this Authority in computer readable form.
   - ☐ The statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.
   - ☐ The statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished.

4. The amendments have resulted in the cancellation of:

   - ☐ the description, pages:
   - ☐ the claims, Nos.:

☐ the drawings, sheets:

5. ☐ This report has been established as if (some of) the amendments had not been made, since they have been considered to go beyond the disclosure as filed (Rule 70.2(c)):

   (Any replacement sheet containing such amendments must be referred to under item 1 and annexed to this report.)

6. Additional observations, if necessary:

V. Reasoned statement under Article 35(2) with regard to novelty, inventive step or industrial applicability; citations and explanations supporting such statement

1. Statement

Novelty (N)	Yes:	Claims	
	No:	Claims	1,16,24
Inventive step (IS)	Yes:	Claims	
	No:	Claims	2-15,17-23,25-30
Industrial applicability (IA)	Yes:	Claims	1-30
	No:	Claims	

2. Citations and explanations
   see separate sheet

VII. Certain defects in the international application

The following defects in the form or contents of the international application have been noted:
see separate sheet

VIII. Certain observations on the international application

The following observations on the clarity of the claims, description, and drawings or on the question whether the claims are fully supported by the description, are made:
see separate sheet

Regarding Point V

(1) EP-A-0 606 646

1. Document (1) represents the state of the art which is closest to the subject-matter of claim 1. The reference teaches a filter material for removing leukocytes from a whole blood product or a red cell product, comprising at least two layers of a filter element which has, in a surface portion thereof, basic functional groups and nonionic hydrophilic groups, wherein a molar ratio of the basic functional groups to the nonionic hydrophilic groups is in the range from 0.6 to 6. The basic functional groups include amino groups and the nonionic hydrophilic groups include a hydroxyl group. Independent claim 1 lacks novelty under PCT Article 33(2) in view of the subject matter disclosed in document (1), as in the filter material of present claim 1, it is not required that the first and second filter elements are different, i.e. that they have on their surface either nitrogen containing functional groups or hydroxyl groups and not both (see also present claim 8). However if novelty can be demonstrated in respect of document (1), an inventive step could be recognised if it could be shown that the specific combination of features as claimed in Claims 1 leads to unexpected advantages in comparison to the filter as disclosed in document (1). Independent Claims 16 and 24 lack novelty under PCT Article 33(2) in view of the subject matter disclosed in document (1). The reference teaches a filter which clearly falls under the scope of Claim 1 and which can be used in the filter device of claim 16 and in the methods for processing a biological fluid of claim 24. Further, no surprising features, whose inception would have involved the exercise of some inventive skill, appear to be present in the dependent claims.

Regarding Point VII

1. The general statement (i.e. "scope and spirit of the invention") in the description at page 26 is not clear, and when used to interpret the claims renders them also unclear, contrary to Article 6 PCT.

2. Reference should not be made to unpublished literature (see Page 1, US

provisional patent application).

3. The description should be made consistent with the claims and should include a proper acknowledgement of the cited prior art; Rule 5.1(a)(ii) PCT.

Regarding Point VIII

1. The dependency of current claim 8 is given incorrectly, claim 8 should be dependent on claim 7.

2. From the wording of the present claim 1 it is not clear which basis should be used for the calculation of the nitrogen-oxygen ratio.

3. The use of the word "about" to qualify the various ranges in the claims is considered to lead to a lack of clarity.

WHAT IS CLAIMED IS:

1. A filter (10) for processing a biological fluid comprising:

at least two filter elements (1, 2), wherein the surface of one filter element (1) has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00, and the surface of the other filter element (2) is hydroxylated relative to the bulk of the element.

2. The filter of claim 1, further comprising at least one additional filter element (1), wherein the surface of the additional element has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00.

3. The filter of claim 1, further comprising at least one additional filter element (2), wherein the surface of the additional element is hydroxylated relative to the bulk of the element.

4. The filter of claim 1, further comprising at least two additional filter elements (1, 2), wherein the surface of the first additional element (1) has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00, and the surface of the second additional element (2) is hydroxylated relative to the bulk of the element.

5. The filter of claim 2, wherein the element (2) having the hydroxylated surface is interposed between the two elements (1, 1) having surfaces including the nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00.

6. The filter of claim 3, wherein the element (1) having a surface including the nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00 is interposed between the two elements (2, 2) having hydroxylated surfaces.

7. The filter of claim 1, wherein at least a portion of the surface of the element (2) hydroxylated relative to the bulk of the element is aminated relative to the bulk of the element.

8. The filter of claim 1, wherein another portion of the surface of the element (2)

hydroxylated relative to the bulk of the element is aminated relative to the bulk of the element.

9. The filter of claim 1, wherein the surface of the filter element (1) has a nitrogen-to-oxygen ratio in the range from at least about 0.2 to less than about 1.00.

10. The filter of claim 1, wherein the filter element (2) with the hydroxylated surface includes at least one carboxyl group.

11. The filter of claim 1, wherein the filter elements (1, 2) have a negative zeta potential at physiological pH.

12. The filter of any one of claims 1 and 7-11, wherein the filter element (1) having the surface including the nitrogen-to-oxygen ratio comprises a porous fibrous leukocyte depletion medium having a first predetermined critical wetting surface tension (CWST); and the filter element (2) having a hydroxylated surface comprises a porous fibrous leukocyte depletion medium having a second predetermined CWST.

13. The filter of claim 12, wherein the two filter elements have different critical wetting surface tensions (CWSTs).

14. The filter of any one of claims 1-13, wherein at least one filter element comprises at least two layers.

15. The filter of any one of claims 1-14, wherein at least one filter element has a CWST of at least about 90 dynes/cm.

16. A filter device (100) for processing a biological fluid comprising:

a housing (25) having an inlet (20) and an outlet (30) and defining a fluid flow path between the inlet and the outlet; and

the filter of any one of claims 1-15 disposed in the housing across the fluid flow

path.

17. The filter device of claim 16, wherein the filter is arranged to allow plasma to pass therethrough and substantially prevent the passage of leukocytes and platelets therethrough.

18. The filter device of claim 16, wherein the filter is arranged to allow plasma to pass therethrough and substantially prevent the passage of leukocytes therethrough, without substantially activating C3a in the biological fluid.

19. The filter device of claim 16, wherein the filter is arranged to allow plasma to pass therethrough and substantially prevent the passage of platelets, leukocytes, and C3a therethrough.

20. The filter device of any one of claims 16-19, wherein the filter is arranged to provide leukocyte-depleted plasma having about 1×10^3 leukocytes or less therein.

21. The filter device of any one of claims 16-20, wherein the filter is arranged to provide platelet-depleted plasma having about 1×10^9 platelets or less therein.

22. The filter device of any one of claims 16-21, wherein the filter substantially removes C3a from the biological fluid passing therethrough.

23. The filter device of any one of claims 16-22, wherein C3a is not substantially activated by the filter as the biological fluid passes through the filter.

24. A method for processing a biological fluid comprising:

passing a biological fluid through the filter device of any one of claims 16-23; and obtaining the filtered fluid.

25. A method for processing a biological fluid comprising:

passing a leukocyte-containing plasma-rich fluid through a filter (10) comprising at

least two filter lements (1, 2), wherein the surface of one filter element (1) has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00, and the surface of the other filter element (2) is hydroxylated relative to the bulk of the element; and

obtaining a filtered plasma-rich biological fluid substantially free of leukocytes and platelets.

26. The method for processing a biological fluid according to claim 25, wherein passing the leukocyte-containing plasma-rich biological fluid through the filter comprising passing the fluid through at least one additional filter element (2), wherein at least a portion of the surface of the element is aminated relative to the bulk of the element, and another portion of the surface of the element is hydroxylated relative to the bulk of the element.

27. The method for processing biological fluid according to claim 25, wherein passing the leukocyte-containing plasma-rich fluid through the filter comprises passing the fluid through at least two additional filter elements (1, 2), the surface of one additional element (1) having a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00, and the surface of the other additional element (2) being hydroxylated relative to the bulk of the element.

28. The method of any one of claims 25-27 wherein the filtered plasma-rich fluid is substantially free of C3a.

29. The method of any one of claims 25-28 wherein the leukocyte-containing plasma-rich biological fluid comprises a platelet-poor biological fluid.

30. The method of any one of claims 25-29, including collecting plasma-rich fluid in a downstream container without substantially activating C3a in the plasma-rich fluid.

PATENT COOPERATION TREATY

From the INTERNATIONAL SEARCHING AUTHORITY

PCT

To:
LEYDIG, VOIT & MAYER
Attn. JAY, J.
700 Thirteenth Street, N.W.
Suite 300
Washington, D.C. 20005
UNITED STATES OF AMERICA

AUG -8 2000

NOTIFICATION OF TRANSMITTAL OF THE INTERNATIONAL SEARCH REPORT OR THE DECLARATION

(PCT Rule 44.1)

Applicant's or agent's file reference 440202/PALL	Date of mailing *(day/month/year)* 03/08/2000
International application No. PCT/US 00/ 06829	FOR FURTHER ACTION See paragraphs 1 and 4 b low
Applicant PALL CORPORATION	International filing date *(day/month/year)* 15/03/2000

1. [X] The applicant is hereby notified that the International Search Report has been established and is transmitted herewith.

Filing of amendments and statement under Article 19:
The applicant is entitled, if he so wishes, to amend the claims of the International Application (see Rule 46):

When? The time limit for filing such amendments is normally 2 months from the date of transmittal of the International Search Report; however, for more details, see the notes on the accompanying sheet.

Where? Directly to the International Bureau of WIPO
34, chemin des Colombettes
1211 Geneva 20, Switzerland
Fascimile No.: (41-22) 740.14.35

For more detailed instructions, see the notes on the accompanying sheet.

2. [ ] The applicant is hereby notified that no International Search Report will be established and that the declaration under Article 17(2)(a) to that effect is transmitted herewith.

3. [ ] **With regard to the protest** against payment of (an) additional fee(s) under Rule 40.2, the applicant is notified that:

[ ] the protest together with the decision thereon has been transmitted to the International Bureau together with the applicant's request to forward the texts of both the protest and the decision thereon to the designated Offices.

[ ] no decision has been made yet on the protest; the applicant will be notified as soon as a decision is made.

4. **Further action(s):** The applicant is reminded of the following:

Shortly after **18 months** from the priority date, the international application will be published by the International Bureau. If the applicant wishes to avoid or postpone publication, a notice of withdrawal of the international application, or of the priority claim, must reach the International Bureau as provided in Rules 90*bis*.1 and 90*bis*.3, respectively, before the completion of the technical preparations for international publication.

Within **19 months** from the priority date, a demand for international preliminary examination must be filed if the applicant wishes to postpone the entry into the national phase until 30 months from the priority date (in some Offices even later).

Within **20 months** from th priority date, the applicant must perform the prescribed acts for entry into the national phase before all designat d Offices which have not been elect d in the demand or in a later election within 19 months from the priority date or could not be elected because they ar not bound by Chapter II.

Nam and mailing address of the International Searching Authority European Pat nt Office, P.B. 5818 Patentlaan 2 NL-2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Authorized officer Heike Zoglauer

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Form PCT/ISA/220 (July 1998)

NOTES TO FORM PCT/ISA/220

These Notes are intended to give the basic instructions concerning the filing of amendments under article 19. The Notes are based on the requirements of the Patent Cooperation Treaty, the Regulations and the Administrative Instructions under that Treaty. In case of discrepancy between these Notes and those requirements, the latter are applicable. For more detailed information, see also the PCT Applicant's Guide, a publication of WIPO.

In these Notes, "Article", "Rule", and "Section" refer to the provisions of the PCT, the PCT Regulations and the PCT Administrative Instructions respectively.

INSTRUCTIONS CONCERNING AMENDMENTS UNDER ARTICLE 19

The applicant has, after having received the international search report, one opportunity to amend the claims of the international application. It should however be emphasized that, since all parts of the international application (claims, description and drawings) may be amended during the international preliminary examination procedure, there is usually no need to file amendments of the claims under Article 19 except where, e.g. the applicant wants the latter to be published for the purposes of provisional protection or has another reason for amending the claims before international publication. Furthermore, it should be emphasized that provisional protection is available in some States only.

What parts of the international application may be amended?

Under Article 19, only the claims may be amended.

During the international phase, the claims may also be amended (or further amended) under Article 34 before the International Preliminary Examining Authority. The description and drawings may only be amended under Article 34 before the International Examining Authority.

Upon entry into the national phase, all parts of the international application may be amended under Article 28 or, where applicable, Article 41.

When?

Within 2 months from the date of transmittal of the international search report or 16 months from the priority date, whichever time limit expires later. It should be noted, however, that the amendments will be considered as having been received on time if they are received by the International Bureau after the expiration of the applicable time limit but before the completion of the technical preparations for international publication (Rule 46.1).

Where not to file the amendments?

The amendments may only be filed with the International Bureau and not with the receiving Office or the International Searching Authority (Rule 46.2).

Where a demand for international preliminary examination has been/is filed, see below.

How?

Either by cancelling one or more entire claims, by adding one or more new claims or by amending the text of one or more of the claims as filed.

A replacement sheet must be submitted for each sheet of the claims which, on account of an amendment or amendments, differs from the sheet originally filed.

All the claims appearing on a replacement sheet must be numbered in Arabic numerals. Where a claim is cancelled, no renumbering of the other claims is required. In all cases where claims are renumbered, they must be renumbered consecutively (Administrative Instructions, Section 205(b)).

The amendments must be made in the language in which the international application is to be published.

What documents must/may accompany the amendments?

Letter (Section 205(b)):

The amendments must be submitted with a letter.

The letter will not be published with the international application and the amended claims. It should not be confused with the "Statement under Article 19(1)" (see below, under "Statement under Article 19(1)").

The letter must be in English or French, at the choice of the applicant. However, if the language of the international application is English, the letter must be in English; if the language of the international application is French, the letter must be in French.

The letter must indicate the differences between the claims as filed and the claims as amended. It must, in particular, indicate, in connection with each claim appearing in the international application (it being understood that identical indications concerning several claims may be grouped),whether

(i) the claim is unchanged;

(ii) the claim is cancelled;

(iii) the claim is new;

(iv) the claim replaces one or more claims as filed;

(v) the claim is the result of the division of a claim as filed.

The following examples illustrate the manner in which amendments must be explained in the accompanying letter:

1. [Where originally there were 48 claims and after amendment of some claims there are 51]:
"Claims 1 to 29, 31, 32, 34, 35, 37 to 48 replaced by amended claims bearing the same numbers; claims 30, 33 and 36 unchanged; new claims 49 to 51 added."

2. [Where originally there were 15 claims and after amendment of all claims there are 11]:
"Claims 1 to 15 replaced by amended claims 1 to 11."

3. [Where originally there were 14 claims and the amendments consist in cancelling some claims and in adding new claims]:
"Claims 1 to 6 and 14 unchanged; claims 7 to 13 cancelled; new claims 15, 16 and 17 added." or
"Claims 7 to 13 cancelled; new claims 15, 16 and 17 added; all other claims unchanged."

4. [Where various kinds of amendments are made]:
"Claims 1-10 unchanged; claims 11 to 13, 18 and 19 cancelled; claims 14, 15 and 16 replaced by amended claim 14; claim 17 subdivided into amended claims 15, 16 and 17; new claims 20 and 21 added."

"Statement under article 19(1)" (Rule 46.4)

The amendments may be accompanied by a statement explaining the amendments and indicating any impact that such amendments might have on the description and the drawings (which cannot be amended under Article 19(1)).

The statement will be published with the international application and the amended claims.

It must be in the language in which the international appplication is to be published.

It must be brief, not exceeding 500 words if in English or if translated into English.

It should not be confused with and does not replace the letter indicating the differences between the claims as filed and as amended. It must be filed on a separate sheet and must be identified as such by a heading, preferably by using the words "Statement under Article 19(1)."

It may not contain any disparaging comments on the international search report or the relevance of citations contained in that report. Reference to citations, relevant to a given claim, contained in the international search report may be made only in connection with an amendment of that claim.

Consequence if a demand for international preliminary examination has already been filed

If, at the time of filing any amendments under Article 19, a demand for international preliminary examination has already been submitted, the applicant must preferably, at the same time of filing the amendments with the International Bureau, also file a copy of such amendments with the International Preliminary Examining Authority (see Rule 62.2(a), first sentence).

Consequence with regard to translation of the international application for entry into the national phase

The applicant's attention is drawn to the fact that, where upon entry into the national phase, a translation of the claims as amended under Article 19 may have to be furnished to the designated/elected Offices, instead of, or in addition to, the translation of the claims as filed.

For further details on the requirements of each designated/elected Office, see Volume II of the PCT Applicant's Guide.

PATENT COOPERATION TREATY

PCT

INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference	FOR FURTHER ACTION	see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, item 5 below.
440202/PALL		
International application No.	International filing date *(day/month/year)*	(Earliest) Priority Date *(day/month/year)*
PCT/US 00/ 06829	15/03/2000	16/03/1999
Applicant		
PALL CORPORATION		

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 3 sheets.

[X] It is also accompanied by a copy of each prior art document cited in this report.

1. **Basis of the report**

   a. With regard to the **language**, the international search was carried out on the basis of the international application in the language in which it was filed, unless otherwise indicated under this item.

   [ ] the international search was carried out on the basis of a translation of the international application furnished to this Authority (Rule 23.1(b)).

   b. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international search was carried out on the basis of the sequence listing :

   [ ] contained in the international application in written form.

   [ ] filed together with the international application in computer readable form.

   [ ] furnished subsequently to this Authority in written form.

   [ ] furnished subsequently to this Authority in computer readble form.

   [ ] the statement that the subsequently furnished written sequence listing does not go beyond the disclosure in the international application as filed has been furnished.

   [ ] the statement that the information recorded in computer readable form is identical to the written sequence listing has been furnished

2. [ ] **Certain claims were found unsearchable** (See Box I).

3. [ ] **Unity of invention is lacking** (see Box II).

4. With regard to the **title**,

   [X] the text is approved as submitted by the applicant.

   [ ] the text has been established by this Authority to read as follows:

5. With regard to the **abstract**,

   [X] the text is approved as submitted by the applicant.

   [ ] the text has been established, according to Rule 38.2(b), by this Authority as it appears in Box III. The applicant may, within one month from the dat of mailing of this international search report, submit comments to this Authority.

6. The figure of the **drawing** to be published with the abstract is Figure No. 1

   [ ] as suggested by the applicant.

   [ ] None of the figures.

   [ ] because the applicant failed to suggest a figure.

   [X] because this figure better characterizes the invention.

PATENT COOPERATION TREA

PCT

INTERNATIONAL SEARCH REPORT

(PCT Article 18 and Rules 43 and 44)

Applicant's or agent's file reference	**FOR FURTHER ACTION** see Notification of Transmittal of International Search Report (Form PCT/ISA/220) as well as, where applicable, item 5 below.	
440202/PALL		
International application No.	International filing date *(day/month/year)*	(Earliest) Priority Date *(day/month/year)*
PCT/US 00/ 06829	15/03/2000	16/03/1999
Applicant		
PALL CORPORATION		

This International Search Report has been prepared by this International Searching Authority and is transmitted to the applicant according to Article 18. A copy is being transmitted to the International Bureau.

This International Search Report consists of a total of 3 sheets.

[X] It is also accompanied by a copy of each prior art document cited in this report.

1. **Basis of the report**

   a. With regard to the **language,** the international search was carried out on the basis of the international application in the language in which it was filed, unless otherwise indicated under this item.

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   b. With regard to any **nucleotide and/or amino acid sequence** disclosed in the international application, the international search was carried out on the basis of the sequence listing :

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2. [ ] **Certain claims were found unsearchable** (See Box I).

3. [ ] **Unity of invention is lacking** (see Box II).

4. With regard to the **title,**

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6. The figure of the **drawings** to be published with the abstract is Figure No. 1

   [ ] as suggested by the applicant.

   [ ] because the applicant failed to suggest a figure.

   [X] because this figure better characterizes the invention.

   [ ] None of the figures.

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 00/06829

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 B01D39/16 A61K35/14 A61K35/16

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 B01D A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)
EPO-Internal

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP 0 606 646 A (ASAHI MEDICAL CO) 20 July 1994 (1994-07-20) claims 7,12 ---	1,2,8, 27,31
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X	EP 0 267 286 A (ASAHI MEDICAL CO) 18 May 1988 (1988-05-18) claim 1 ---	7,8,27
X	EP 0 630 675 A (TERUMO CORP) 28 December 1994 (1994-12-28) page 4, line 48 -page 5, line 10 --- -/--	30,32

[X] Further documents are listed in the continuation of box C.

[X] Patent family members are listed in annex.

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Date of the actual completion of the international search	Date of mailing of the international search report
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INTERNATIONAL SEARCH REPORT

International Application No
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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication,where appropriate, of the relevant passages	Relevant to claim No.
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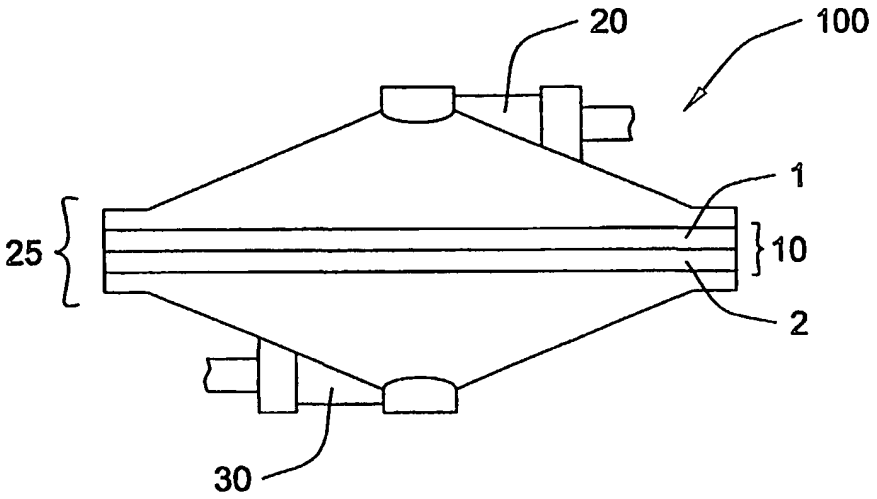

(57) Abstract

A filter device (100) comprising a filter (10) for producing a leukocyte–depleted plasma–rich fluid is disclosed.

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BIOLOGICAL FLUID FILTER AND SYSTEM

This application claims the benefit of U.S. provisional patent application 60/124,580, filed March 16, 1999, which is incorporated by reference.

TECHNICAL FIELD

This invention relates to a filter for processing a biological fluid, more particularly, a filter that provides a leukocyte-depleted, substantially platelet-free biological fluid. Preferably, the filter provides a biological fluid that is substantially free of platelets, leukocytes, and biologically active complement fragments.

BACKGROUND OF THE INVENTION

Vertebrate blood contains a number of components, including plasma, platelets, and red blood cells. Blood also contains components such as various types of white blood cells (leukocytes), and proteins of the complement system, that provide for combating infection.

Blood components may be separated, and further processed, for a variety of uses, particularly as transfusion products. Illustratively, red blood cells (typically concentrated as packed red blood cells), plasma, and platelets (typically concentrated as platelet concentrate), can be separately administered to different patients. Some components, e.g., plasma and/or platelets, can be pooled before administration, and plasma can be fractionated to provide enriched components to treat disease.

While leukocytes combat infection and engulf and digest invading microorganisms and debris, the presence of leukocytes in transfusion products can be undesirable, since, for example, they may cause adverse effects (e.g., a febrile reaction) in the patient receiving the transfusion. Additionally, the presence of a significant level of red blood cells in some transfusion products (particularly if the transfusion products have been pooled) can lead to an adverse immune response by the patient.

The processing of blood to produce transfusion products can lead to the activation of the complement system, that acts on its own and in cooperation with

antibodies in defending the host against infection. The complement system is composed of a series of plasma-borne blood proteins (proenzymes) that are sequentially activated in a series of reactions. The proteins are activated in cascade fashion, i.e., the output of one reaction is the input for the next. The cascade ultimately generates a terminal five-protein membrane attack complex (MAC, C5b-9), whose physiological function is protection of the host from invading microorganisms. The MAC causes lysis of the microorganisms.

While the complement system is generally beneficial in protecting the host, the presence of the various activated or activatable blood proteins (and fragments thereof) can be undesirable, particularly when these proteins and/or fragments are present in blood or blood components used for transfusion. For example, activation can lead to the administration of biologically active complement fragments such as C3a and its metabolite, C3a des Arg^{77}. Transfusing activated complement into a patient can cause adverse affects such as anaphylactoid reactions, platelet aggregation, and/or immune suppression.

Accordingly, there is a need in the art for a filter for use with biological fluids such as blood and blood components, particularly for the production of plasma-rich blood products, that minimizes the contamination of the plasma-rich blood product by leukocytes, as well by other materials such as platelets and/or red blood cells. There is also a need for a filter that depletes complement and/or prevents the activation of complement in biological fluids such as plasma-rich blood products. These and other advantages of the present invention will be apparent from the description as set forth below.

SUMMARY OF THE INVENTION

In accordance with an embodiment of the invention, a filter device for providing a plasma-rich biological fluid substantially free of leukocytes comprises a filter including a first filter element and a second filter element, wherein the first filter element comprises a porous fibrous leukocyte depletion medium having a first predetermined critical wetting surface tension (CWST), and the second filter element, arranged

downstream of the first filter element, comprises a porous fibrous leukocyte depletion medium having a second predetermined CWST. Typically, the CWST of the first element differs from the CWST of the second element.

In accordance with another embodiment of the invention, a filter for processing a biological fluid is provided comprising at least one filter element, wherein at least a portion of the surface of the element is aminated and hydroxylated relative to its bulk, or a portion of the surface of the element is aminated, and another portion of the surface of the element is hydroxylated, relative to the element's bulk.

In some embodiments, a filter according to the invention comprises at least two filter elements, wherein the surface of one filter element has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00 and/or the surface of the element is aminated relative to its bulk, and the surface of the other filter element is hydroxylated relative to its bulk. In one embodiment, one filter element has a CWST that differs from the CWST of the other element.

In preferred embodiments, the filter allows plasma-rich biological fluid to pass therethrough and substantially prevent the passage of leukocytes and platelets. In some embodiments, the filter substantially removes at least one biologically active complement fragment such as C3a and/or the filter does not substantially activate the fragment(s).

Methods for using the filter, the filter device, and systems including the filter device are also provided.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is an embodiment of a filter device according to the present invention.

Figure 2 is another embodiment of a filter device according to the present invention.

Figure 3 is another embodiment of a filter device according to the present invention.

Figure 4 is an embodiment of a biological fluid processing system including a filter device according to the invention.

Figure 5 (A and B) illustrates other embodiments of biological fluid processing systems including a filter device according to the invention.

Figure 6 is another embodiment of a biological fluid processing system including a filter device according to the invention.

SPECIFIC DESCRIPTION OF THE INVENTION

In accordance with an embodiment of the present invention, a filter device for processing a biological fluid comprises a housing having an inlet and an outlet and defining a fluid flow path between the inlet and the outlet, a filter disposed in the housing across the fluid flow path, the filter comprising at least one filter element comprising a porous fibrous leukocyte depletion medium having a first predetermined critical wetting surface tension (CWST), and at least one filter element comprising a porous fibrous leukocyte and platelet depletion medium having a second predetermined CWST, wherein the filter is arranged to allow plasma to pass therethrough and substantially prevent the passage of leukocytes therethrough.

A filter device for processing a biological fluid in accordance with another embodiment of the invention comprises a housing having an inlet and an outlet and defining a fluid flow path between the inlet and the outlet, a filter disposed in the housing across the fluid flow path, the filter comprising at least one filter element comprising a porous fibrous leukocyte depletion medium having a first predetermined CWST, and at least one filter element comprising a porous fibrous leukocyte and platelet depletion medium having a second predetermined CWST, wherein the filter is arranged to allow plasma to pass therethrough and substantially prevent the passage of leukocytes therethrough, without substantially activating C3a in the biological fluid.

In accordance with another embodiment of the invention, a filter device for processing a biological fluid comprises a housing having an inlet and an outlet and defining a fluid flow path between the inlet and the outlet, a filter disposed in the housing across the fluid flow path, the filter comprising at least one filter element comprising a porous fibrous leukocyte depletion medium having a first predetermined CWST, and at least one filter element comprising a porous fibrous leukocyte and platelet

depletion medium having a second predetermined CWST, wherein the filter is arranged to allow plasma to pass therethrough and substantially prevent the passage of platelets, leukocytes and C3a therethrough.

In another embodiment, a filter for processing a biological fluid comprises at least one filter element, preferably a fibrous element, wherein at least a portion of the surface of the element is aminated and hydroxylated relative to its bulk, or a portion of the surface of the element is aminated, and another portion of the surface of the element is hydroxylated, relative to the element's bulk.

In some embodiments, a filter according to the invention comprises at least two filter elements, wherein the surface of one filter element has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00 and/or the surface of the element is aminated relative to its bulk, and the surface of the other filter element is hydroxylated relative to its bulk.

Preferably, the filter includes at least one additional filter element, the filter element comprising any of the elements described above.

Embodiments providing leukocyte-depleted, substantially platelet-free biological fluid can be especially desirable, as potential disease-causing agents such as prions (implicated as causing disease, e.g., degenerative diseases such as Creutzfeld-Jacob disease (CJD) and "mad cow" disease) may attach to platelets and/or leukocytes, and the attached agents would be removed (and thus not transmitted to the patient during transfusion) upon removal of the leukocytes and the platelets.

A method for processing a biological fluid in accordance with an embodiment of the invention comprises passing a leukocyte-containing plasma-rich biological fluid through a filter device comprising a filter including a first element comprising a porous fibrous leukocyte depletion medium having a first predetermined CWST, and a second element comprising a porous fibrous leukocyte depletion medium having a second predetermined CWST, and collecting a filtered plasma-rich biological fluid substantially free of leukocytes.

A method for processing a biological fluid in accordance with another embodiment comprises passing a leukocyte-containing plasma-rich biological fluid

through a filter device comprising a filter including at least one filter element, wherein at least a portion of the surface of the element is aminated and hydroxylated relative to its bulk, or a portion of the surface of the element is aminated, and another portion of the surface of the element is hydroxylated, relative to the element's bulk, and obtaining a filtered plasma-rich biological fluid substantially free of leukocytes and platelets.

In accordance with another embodiment, a method for processing a biological fluid comprises passing a leukocyte-containing plasma-rich biological fluid through a filter device comprising a filter including at least two filter elements, wherein the surface of one filter element has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00 and/or the surface of the element is aminated relative to its bulk, and the surface of the other filter element is hydroxylated relative to its bulk, and obtaining a filtered plasma-rich biological fluid substantially free of leukocytes and platelets.

In some embodiments, the method includes passing biological fluid through the filter device without substantially activating complement in the biological fluid. For example, the level of the biologically active complement fragment C3a in the filtered fluid is not substantially increased as compared to the level in the fluid before filtration. In an embodiment, the method includes depleting the fluid of a biologically active complement fragment (e.g., C3a) upon passing it through the filter device.

Another embodiment of a method for processing a biological fluid comprises passing a leukocyte-containing plasma-rich biological fluid through a filter device comprising a filter including a fibrous leukocyte depletion medium and a fibrous leukocyte and platelet depletion medium, and collecting a filtered plasma-rich biological fluid substantially free of platelets and leukocytes.

A method for processing a biological fluid according to another embodiment comprises passing a leukocyte-containing plasma-rich biological fluid through a filter device comprising a filter including a fibrous leukocyte depletion medium and a fibrous leukocyte and platelet depletion medium, and collecting a filtered plasma-rich biological fluid substantially free of leukocytes, and C3a.

The invention also provides a method for processing a biological fluid comprising passing a leukocyte-containing plasma-rich biological fluid through a filter

device comprising a filter including a fibrous leukocyte depletion medium and a fibrous leukocyte and platelet depletion medium, wherein the filter is arranged to substantially prevent the passage of red blood cells therethrough, and collecting a filtered plasma-rich biological fluid substantially free of platelets, leukocytes, and red blood cells.

In accordance with yet another embodiment of the invention, a method for processing a biological fluid is provided comprising passing a platelet-poor plasma-rich biological fluid through a filter device comprising a filter including a fibrous leukocyte depletion medium and a fibrous leukocyte and platelet depletion medium, and collecting a filtered plasma-rich biological fluid substantially free of platelets and leukocytes.

In some embodiments of the method, a filtered substantially cell-free plasma-containing fluid is provided, wherein the fluid is substantially free of C3a.

A system according to an embodiment of the invention comprises a filter device, and at least one container such as a plastic blood bag, in fluid communication with the filter device. Typical embodiments of the system include a filter device, interposed between, and in fluid communication with, at least two containers such as plastic blood bags. In one preferred embodiment, the system comprises a closed system.

As used herein, the term "complement" includes at least one of a complement protein, complement component (e.g., C1 through C9), complement fragment, biologically active fragment of a component (and metabolite of the fragment), complement factor (e.g., factor B and factor D), complement subcomponent, and complement complex (e.g., $\overline{C567}$). Exemplary biologically active fragments and metabolites thereof include C3a, C3a des Arg^{77}, C4a, C4a des Arg, C5a, and C5a des Arg.

As used herein, a biological fluid includes any treated or untreated fluid associated with living organisms, particularly blood, including whole blood, warm or cold blood, and stored or fresh blood; treated blood, such as blood diluted with at least one physiological solution, including but not limited to saline, nutrient, and/or anticoagulant solutions; blood components, such as platelet concentrate (PC), platelet-rich plasma (PRP), platelet-poor plasma (PPP), platelet-free plasma, plasma, components obtained from plasma, packed red cells (PRC), transition zone material or buffy coat (BC); blood products derived from blood or a blood component or derived

from bone marrow; red cells separated from plasma and resuspended in a physiological fluid or a cryoprotective fluid; and platelets separated from plasma and resuspended in a physiological fluid or a cryoprotective fluid. The biological fluid may have been treated to remove some of the leukocytes before being processed according to the invention. As used herein, blood product or biological fluid refers to the components described above, and to similar blood products or biological fluids obtained by other means and with similar properties.

A "unit" is the quantity of biological fluid from a donor or derived from one unit of whole blood. It may also refer to the quantity drawn during a single donation. Typically, the volume of a unit varies, the amount differing from donation to donation. Multiple units of some blood components, particularly platelets and buffy coat, may be pooled or combined, typically by combining four or more units.

As used herein, the term "closed" refers to a system that allows the collection and processing (and, if desired, the manipulation, e.g., separation of portions, separation into components, filtration, storage, and preservation) of biological fluid, e.g., donor blood, blood samples, and/or blood components, without the need to compromise the integrity of the system. A closed system can be as originally made, or result from the connection of system components using what are known as "sterile docking" devices. Illustrative sterile docking devices are disclosed in U.S. Patent Nos. 4,507,119, 4,737,214, and 4,913,756.

Each of the components of the invention will now be described in more detail below, wherein like components have like reference numbers.

In accordance with the invention, a filter device comprises a housing having an inlet and an outlet, and defining a fluid flow path between the inlet and the outlet, wherein a filter comprising at least one filter element is disposed across the fluid flow path. Figures 1-3 illustrated several embodiment of the filter device 100, comprising a housing 25 having an inlet 20 and an outlet 30, and defining the fluid flow path between the inlet and the outlet, wherein a filter 10, comprising at least one first filter element 1 and at least one second filter element 2, is disposed across the fluid flow path. Figure 1 illustrates a filter 10 comprising a first filter element 1 and a second filter element 2,

while Figures 2 and 3 illustrate filters having a plurality of first filter elements or second filter elements. In other embodiments (not shown) the filter comprises at least one filter element.

In accordance with the invention, the filter 10 is arranged to prevent the passage therethrough of an undesirable level of leukocytes, and typically, to prevent the passage of an undesirable level of platelets. In a preferred embodiment, the filter is arranged to prevent the passage therethrough of an undesirable level of red blood cells. Even more preferably, the filter is arranged to substantially remove at least one biologically active complement fragment such as C3a, and/or to substantially minimize activation of the complement fragment.

The filter 10 may be configured to remove a desired amount of leukocytes. Typically, the filter is configured to remove greater than about 90%, preferably, greater than about 99%, even more preferably, greater than about 99.9%, or more, of the leukocytes from the plasma-rich fluid passing through the filter. For example, the filter can be configured to provide a filtered fluid having about 1×10^4 leukocytes or less. In some embodiments, the filter can be configured to provide a filtered fluid having about 1×10^3 leukocytes or less. In one embodiment, wherein the fluid to be filtered comprises platelet-poor-plasma, the resultant filtered fluid has about 200 leukocytes/liter or less, preferably, about 100 leukocytes/liter or less. In some embodiments, the filtered fluid has about 75 leukocytes/unit (e.g., wherein a unit has volume of about 300 ml) or less.

Typically, the filter is configured to prevent the passage therethrough of a significant level of platelets, and can be configured to prevent the passage therethrough of a substantial number of red blood cells. The filter can also be configured to reduce the passage therethrough of blood cell fragments.

Illustratively, the filtered fluid (e.g., in the container downstream of the filter) preferably contains less than about 5000 platelets/μL. For example, the resultant unit of filtered fluid can have less than about 1×10^9 platelets. In one preferred embodiment, the resultant unit of filtered fluid has about 1×10^4 platelets, or less, or about 1×10^3 platelets, or less. Preferably, there should be no visible indication (to the technician carrying out the filtration) of red blood cells downstream of the filter.

Preferably, the filter substantially minimizes activation of at least one complement fragment, e.g., a biologically active complement fragment such as C3a, in the biological fluid and/or removes at least one complement fragment from the biological fluid.

With respect to minimizing activation of at least one complement fragment, in an embodiment, the filtered fluid has a C3a level substantially similar to the C3a level in the unfiltered fluid, e.g., of about 900 ng/ml or less, in some embodiments, about 750 ng/ml or less.

With respect to depleting complement, the fluid can be filtered to provide a desired level of complement depletion. For example, in an embodiment, a biological fluid having a C3a level in the range of about 750 to about 900 ng/ml can be filtered to provide a fluid having a C3a level of about 500 ng/ml or less, in some embodiments, about 250 ng/ml or less.

The filter is arranged to filter a suitable volume of fluid in a suitable amount of time. For example, the filter can be capable of filtering about 200 to about 1000 ml of fluid without a significant effect on the overall processing time.

Illustratively, in some embodiments, the filter is capable of filtering about 250 to about 350 ml of fluid in about 15 minutes, or less, e.g., in about 10 minutes or less. In one embodiment, the filter is capable of filtering about 250 to about 350 ml of fluid in about 6 minutes.

In some other embodiments, the filter is capable of filtering about 500 to about 1000 ml of fluid in about 25 minutes, or less, preferably, about 20 minutes or less. In one embodiment, the filter is capable of filtering about 600 to about 850 ml of fluid (e.g., a unit of apheresed plasma) in about 18 minutes or less.

Preferably, one or more filter elements, e.g., each of the first element and the second element 1, 2 of the filter 10, typically comprising depth filter elements, comprise leukocyte depletion media, wherein at least some of the leukocytes are removed by adsorption. In some embodiments, the element, or the first and/or second element ,also remove at least some of the leukocytes by filtration (e.g., sieving). If desired, at least one element removes red blood cells by filtration.

A variety of materials can be used, including synthetic polymeric materials, to produce the porous media of the filter elements according to the invention. Suitable synthetic polymeric materials include, for example, polyolefins, polyesters, and polyamides. Illustrative suitable materials include polybutylene terephthalate (PBT), polyethylene, polyethylene terephthalate (PET), polypropylene, polymethylpentene, polyvinylidene fluoride, polyethersulfone, and nylon, such as nylon 6, nylon 66, nylon 612, nylon 11, and nylon 6 copolymers.

In one preferred embodiment, at least one element, e.g., the first element 1, and the second element 2, each comprise a fibrous medium, typically a fibrous polymeric medium prepared from melt-blown fibers, as disclosed in, for example, U.S. Patent Nos. 4,880,548; 4,925,572, 5,152,905, 5,258,127, 5,443,743, and 5,472,621, as well as International Publication Nos. WO 91/04088, WO 93/04763, and WO 96/03194. An element, which can comprise a preformed medium, can include a plurality of layers and/or media.

One or more elements, e.g., first element 1 and/or the second element 2, can be treated for increased efficiency in processing a biological fluid. For example, surface characteristics of the first and/or second element can be modified (e.g., to affect the CWST, to include a surface charge, e.g., a positive or negative charge, and/or to alter the polarity or hydrophilicity of the surface) by chemical reaction including, for example, wet or dry oxidation, by coating or depositing a polymer on the surface, or by a grafting reaction. Modifications include, e.g., irradiation, a polar or charged monomer, coating and/or curing the surface with a charged polymer, and carrying out chemical modification to attach functional groups on the surface. Grafting reactions may be activated by exposure to an energy source such as gas plasma, heat, a Van der Graff generator, ultraviolet light, electron beam, or to various other forms of radiation, or by surface etching or deposition using a plasma treatment. The materials used to produce the elements can be treated before fabricating the elements, or the elements can be fabricated and subsequently treated.

Illustratively, any one or more of the elements may be surface modified to affect the critical wetting surface tension (CWST), as described in, for example, the U.S.

Patents and International Publications listed above.

Typically, the first element 1 and the second element 2 each have a CWST of at least about 55 dynes/cm, more typically, at least 58 dynes/cm. Preferably, the first element 1 and the second element 2 each have a CWST of at least about 62 dynes/cm. The CWST of one element (e.g., the first element) can be different than the CWST of another element (e.g., the second element). Illustratively, one of the elements can have a CWST in the range of from about 58 dynes/cm to about 75 dynes/cm, and another element can have a CWST in the range of from about 78 dynes/cm to about 115 dynes/cm.

Preferably, at least one of the elements (e.g., the first element or the second element) has a CWST of greater than about 70 dynes/cm. For example, the element may have a CWST in the range from about 75 dynes/cm to about 115 dynes/cm, e.g., in the range of about 80 to about 100 dynes/cm. In some embodiments, the element has a CWST of about 85 dynes/cm, or greater, e.g., in the range from about 90 to about 105 dynes/cm, or in the range from about 85 dynes/cm to about 98 dynes/cm.

In those embodiments including a plurality of elements, while both the first and second elements preferably comprise leukocyte depletion media, the elements typically differ with respect to levels or efficiencies of platelet removal, as well as complement removal and/or inactivation.

For example, the first element typically comprises a platelet depletion medium, and the second element typically removes biologically active complement fragments such as C3a and/or does not substantially activate such biologically active complement fragments. Typically, as will be described in more detail below, the surface of the first element has a nitrogen-to-oxygen ratio in the range of from at least about 0.01 to less than about 1.00, and the second element has an hydroxylated surface.

In some embodiments, the first element (comprising a platelet depletion medium) comprises a medium that has been surface modified by exposure to gas plasma. The plasma can be generated by any suitable method, preferably, by electrical discharge, e.g., radio frequency (RF) discharge. The gas used to treat the surface of the medium can include one or more inorganic and/or organic gases. Illustrative inorganic gases include

helium, argon, nitrogen, neon, nitrous oxide, nitrogen dioxide, oxygen, air, ammonia, carbon monoxide, carbon dioxide, hydrogen, chlorine, hydrogen chloride, bromine cyanide, sulfur dioxide, hydrogen sulfide, xenon, krypton, and the like. Illustrative organic gases include acetylene, pyridine, gases of organosilane compounds and organopolysiloxane compounds, fluorocarbon compounds, and the like. In addition, the gas can be a vaporized organic material, such as an ethylenic monomer to be plasma polymerized or deposited on the surface of the medium. These gases may be used either singly, or as a mixture of two kinds (e.g., two inorganic gases, two organic gases, and inorganic gas and an organic gas) or more. For example, the atmosphere where plasma is generated can includes a carrier gas, e.g., helium or argon.

In some preferred embodiments, the medium is exposed to a gas plasma generated in an atmosphere comprising a nitrogen-containing molecule to obtain a plasma treated substrate. Any suitable nitrogen-containing molecule can be used, one preferred nitrogen-containing molecule is ammonia. As noted above, the atmosphere where plasma is generated can include a carrier gas.

Examples of other nitrogen-containing molecules include alkylamines, allylamines, alkylimines, ethanolamines, hydroxylamines, nitro compounds such as, for example, nitroalkanes, and amides such as, for example, formamide and acetamide.

In one preferred embodiment, surface of the first element has a nitrogen-to-oxygen ratio in the range of from at least about 0.01 to less than about 1.00. In a more preferred embodiment, the nitrogen-to-oxygen ratio is in the range of from at least about 0.02 to less than 1.00. In some embodiments, the surface of the first element is substantially non-hydroxylated, e.g., having less than about 0.1% hydroxyl groups (less than about 1000 ppm).

Alternatively, or additionally, in an embodiment, the first element has a surface characterized by one or more, and in some embodiments, two or more, of the following: a surface that is hydroxylated (has more hydroxyl groups) relative to its bulk, i.e., the surface is more hydroxylated than the interior portion of the element adjacent or under the surface and/or is more hydroxylated than the substrate of the element; a surface that is aminated (has more amine groups) relative to its bulk; a surface that has a greater

number of carbonyl groups relative to its bulk; a surface that has a greater number of carboxyl groups relative to its bulk; a surface that has more ether groups relative to its bulk; and a surface that has more amido groups relative to its bulk.

Illustrative gases and gas plasma treatments include those disclosed in, for example, U.S. Patent Nos. 5,258,127, 5,443,743, 5,679,264, as well as in International Publication Nos. WO 93/04763 and WO 96/03194.

In some embodiments, the filter element that removes biologically active complement fragments such as C3a and/or does not substantially activate such biologically active complement fragments (i.e., the second element) comprises a medium that has been treated (e.g., surface modified) to include a high density of hydroxyl groups, more preferably, to also include anionic groups, e.g., some carboxyl groups as well as the high density of hydroxyl groups.

For example, the second element can have a hydroxylated surface, and in an embodiment, has a grafted coating comprising hydroxyl groups, e.g., comprising an hydroxylated polymer, such as, but not limited to, an hydroxyl acrylate polymer. In some embodiments including a hydroxylated polymer, the polymer further comprises carboxyl groups, e.g., a copolymer including a hydroxyl-containing monomer and a carboxyl containing monomer, such as, but not limited to, a copolymer of hydroxyalkylacrylate and acrylic acid.

In an exemplary technique, at least one of a variety of monomers each comprising an ethylene or acrylic moiety and a second group, which can be selected from hydrophilic groups (e.g., -COOH, or -OH) are used, e.g., in radiation grafting. Grafting of the medium can also be accomplished by compounds containing an ethylenically unsaturated group, such as an acrylic moiety, combined with a hydroxyl group, e.g., monomers such as hydroxyethyl methacrylate (HEMA) or acrylic acid. The compounds containing an ethylenically unsaturated group may be combined with a second monomer such as methacrylic acid (MAA). In an embodiment, the medium is surface modified using a mixture including hydroxyl-terminated and carboxyl-terminated monomers.

Illustrative compounds and groups, e.g., hydroxyl groups and carboxyl groups, as

well as illustrative medium treatment protocols include, but are not limited to, those disclosed in U.S. Patent Nos. 5,152,905, 4,880,548 and 4,925,572, as well as International Publication No. WO 91/04088.

In some embodiments, one or more elements, and typically, both the first and second filter elements, has a negative zeta potential at physiological pH (e.g., about 7 to about 7.4).

For example, the filter element comprising a platelet depletion medium can have a zeta potential of about –3 millivolts (mv), at physiological pH, or the zeta potential can be more negative, e.g., in the range of from about –5 mv to about –25 mv. In some embodiments, the platelet depletion medium has a zeta potential in the range from about –8 mv to about –20 mv at physiological pH.

The filter element comprising a medium that removes biologically active complement fragments and/or does not substantially activate the fragments can have can have a zeta potential of about –3 millivolts (mv) at physiological pH, or the zeta potential can be more negative, e.g., in the range of from about –5 mv to about –20 mv. In some embodiments, the medium has a zeta potential in the range from about –7 mv to about –15 mv at physiological pH.

In some embodiments wherein both types of media have a negative zeta potential at physiological pH, one medium can have a zeta potential that is more negative than that of the other medium.

Typically, the filter has a pore structure that reduces the passage therethrough of white and/or red blood cells. The pore structure can be characterized in terms of pore size, which may be determined by a variety of techniques known to the ordinary artisan. Illustratively, the pore structure can refer to the pore rating or pore diameter as measured by, for example, the modified F2 test, e.g., as described in U.S. Patent Nos. 4,925,572 and 5,229,012. The pore structure can refer to an average pore size as measured by, for example, a Coulter Instruments porometer (e.g., a Coulter Porometer II® machine). Other suitable techniques for determining pore structure values include bubble point tests and Latex Sphere Tests.

For example, the filter can have a pore diameter of about 8 micrometers (μm) or

less. In another embodiment, the filter and/or at least one filter element has a pore diameter of about 5 micrometers or less. In another embodiment, the filter has a pore diameter of about 2.5 micrometers, or less.

The filter can include a plurality of filter elements having different pore structures and/or at least one element can have a varied pore structure.

Typically, each of the types of media has a voids volume of at least about 70%, more preferably, at least 75%. In one embodiment, each of the types of media has a voids volume of about 80% or more. In some embodiments, each of the types of media has a voids volume in the range of from about 85% to about 96%. Typically, in those embodiments wherein the filter includes two or more elements forming a laminate, the laminate has a voids volume in the range of from about 75% to about 85%.

In accordance with the invention, the filter can include a plurality of first elements and/or second elements, arranged in a variety of configurations. Illustratively, Figures 1-3 show exemplary configurations, wherein at least a portion of the filter 10 has alternating elements. In some embodiments, the first and second elements can alternate, or there can be two or more first elements followed by one or more second elements, and/or other combinations. For example, a portion of the filter can include alternating elements, and at least another portion of the filter can include a plurality of the same type of element.

Either element can be the most upstream (e.g., closest to the inlet of the filter device) or downstream (e.g., closest to the outlet). Alternatively, the most upstream and downstream element can be the same (e.g., the second element), with the other type of element (e.g., the first element) interposed between the upstream and downstream elements.

The filter 10 can include additional elements, layers, or components, that can have different structures and/or functions, e.g., at least one of prefiltration, support, drainage, spacing and cushioning. Illustratively, the filter can also include at least one additional element such as a mesh and/or a screen.

The filter 10, comprising the first and second elements, is typically placed in a housing 25 to form a filter assembly or filter device 100. Preferably, the filter device is

sterilizable. Any housing of suitable shape to provide an inlet and an outlet may be employed. The housing is preferably fabricated from any suitable impervious material, including any impervious thermoplastic material, which is compatible with the fluid being processed. The housing may include an arrangement of one or more channels, grooves, conduits, passages, ribs, or the like, which may be serpentine, parallel, curved, circular, or a variety of other configurations.

Suitable exemplary housings are disclosed in U.S. Patent Nos. 5,100,564, 5,152,905, 4,923,620, 4,880,548, 4,925,572, and 5,660,731, as well as International Publication No. WO 91/04088. It is intended that the present invention not be limited by the type, shape, or construction of the housing.

Typically, the filter device or filter assembly 100 according to the invention is included in a biological fluid processing system, e.g., a system including a plurality of conduits and containers, preferably flexible containers such as blood bags. In one preferred embodiment, a system according to the invention comprises a closed system including the filter device.

Figures 4 and 6 illustrate exemplary embodiments of a biological fluid processing system 1000, including the filter device 100 and a plurality of containers 50-53, wherein the components of the system are in fluid communication with each other via a plurality of conduits and connectors, e.g., conduits 60-79 and 91-96, and connectors 80-84. Typically, the system includes at least one, and more typically, at least two flow control devices such as clamps, valves, and/or transfer leg closures. For example, the embodiment illustrated in Figure 4 includes a flow control device 40 and the embodiment illustrated in Figure 6 includes two flow control devices 40. In a variation of the system illustrated in Figure 4, additional flow control devices such as clamps and/or transfer leg closures are associated with conduits 63, 65, and 67. In a variation of the system illustrated in Figure 6, additional flow control devices such as clamps and/or transfer leg closures are associated with conduits 70, 74, and 72.

In both of these illustrated embodiments, the system 1000 also includes a phlebotomy needle 501 (with a cover), a phlebotomy needle protector 500 (e.g., having at least a flexible side wall and being capable of sliding along a conduit and retaining the

phlebotomy needle therein as described in International Publication No. WO 00/06229), a sampling arrangement 600, a sampling arrangement needle or cannula 601 (with a cover), and an additional filter device, leukocyte filter device 200. In those embodiments including a sampling arrangement 600, the arrangement is preferably disposed to minimize contamination of the collected biological fluid by allowing a first sample or portion of the collected fluid (that may be more susceptible to contamination) to be passed to a location other than the collection container 50 (e.g., as described in International Publication Nos. WO 00/07642 and WO 98/28057). For example, the first sample is passed from phlebotomy needle 501 through the sampling arrangement 600 and sampling arrangement needle 601 into a sampling device (not shown) such as an evacuated stoppered collection device. In other embodiments (not shown) the sampling arrangement does not include a needle or cannula, and allows portions or samples of fluid to be passed into one or more attached containers or reservoirs while maintaining a closed system. After the first portion of fluid is passed through the sampling arrangement, a second portion of fluid (that may be less susceptible to contamination) can be collected in the collection container.

One or more containers in the system can be suitable for holding, for example, blood components and/or additives (e.g., nutrients, storage solutions, and/or inactivation agents). The system can include additional components, such as, for example, additional filter devices, including leukocyte depletion filter devices, (with and without filter bypass loops). Additionally, or alternatively, the system can include at least one of the following: a vent such as a gas collection and displacement arrangement, one or more gas inlets, one or more gas outlets, at least one flow control device such as a clamp, transfer leg closure or valve, as well as a sampling arrangement, one or more needles and/or cannulas, and a phlebotomy needle protector.

In the embodiments of the system 1000 illustrated in Figures 4 and 6, the system includes leukocyte filter device 200, e.g., to reduce the level of leukocytes from a red blood cell-containing biological fluid (e.g., packed red blood cells or whole blood). The red blood cell-containing fluid can be further processed.

For example, in accordance with one embodiment of the system illustrated in

Figure 4, a unit of biological fluid, e.g., a unit of whole blood, is passed through phlebotomy needle 501 along conduits 60 and 62 into collection bag 50, that typically contains an additive such as an anticoagulant. If desired, a first portion of biological fluid can be passed through the sampling arrangement 600 before collecting the unit of biological fluid. For example, a first portion of blood can be passed through phlebotomy needle 501, and along conduit 60, connector 80, conduit 61, and through needle 601 into a sampling device. Alternatively, the sampling arrangement can include at least one container or reservoir rather than a needle, and the portion(s) can be passed into the container or reservoir. In another embodiment, the portion of blood is passed through the sampling arrangement after collecting the unit of biological fluid.

Continuing to use the illustrative system shown in Figure 4 for reference, the unit of biological fluid is typically centrifuged to provide a supernatant layer comprising a plasma-rich fluid (e.g., platelet-poor-plasma), a sediment layer comprising red blood cells, and a buffy coat layer between the supernatant and sediment layers. Alternatively, the unit of blood is centrifuged to provide a supernatant layer comprising plasma-rich fluid, and a sediment layer comprising red blood cells.

Subsequently, the plasma-rich fluid (e.g., platelet-poor plasma) is passed from the collection container 50 along conduits 63 and 64 into first satellite container 51 through the filter device 100, i.e., the device comprising a filter 10 having first and second filter elements 1 and 2 as described above, to provide plasma-rich fluid substantially free of leukocytes and platelets without externally visible red blood cells in first satellite container 51. In an embodiment, the filtered plasma-rich fluid is also substantially free of at least one biologically active complement fragment such as C3a.

The separated blood components can be further processed if desired. For example, in accordance with the embodiment of the system illustrated in Figure 4, the sediment layer (comprising red blood cells) can be passed along conduit 65 into second satellite container 52, and subsequently passed along conduit 66, through leukocyte filter device 200 and along conduit 67 into third satellite container 53. In some embodiments, different blood components can be passed essentially simultaneously from separate ports of the collection bag 50, for example, along conduit 63 (e.g., plasma-rich fluid) and

conduit 65 (red blood cell-rich fluid).

If desired, the red blood cells can be combined with an additive solution (before or after filtration). After filtration, the red cells, or the red cells combined with additive solution, can be stored until needed.

In accordance with an embodiment of the system illustrated in Figure 6, a unit of whole blood can be passed through phlebotomy needle 501 along conduits 60 and 69 into collection bag 50, that typically contains an additive such as an anticoagulant. If desired, one or more portions of biological fluid can be passed through the sampling arrangement 600 as described with respect to Figure 4.

The biological fluid is subsequently passed into satellite container 53 along conduit 70, connector 82, and conduit 71, through leukocyte filter device 200 (in some embodiments, a leukocyte- and platelet-depleting filter device that allows red cells and plasma to pass therethrough), along conduit 72, connector 83, and conduit 73. If desired, any embodiments of a system can include additional components such as at least one vent, e.g., a gas inlet and or a gas outlet, or include a bypass loop and/or a gas displacement loop. For example, Figure 6 shows a bypass loop communicating with the filter device 200 (e.g., comprising conduit 79, and a check valve (not shown) between the ends of the loop).

Preferably, the biological fluid in satellite container 50 is centrifuged to provide a supernatant layer comprising plasma-rich fluid (e.g., platelet-poor-plasma) and a sediment layer comprising red blood cells, and the supernatant layer of plasma rich fluid is passed through filter device 100 (i.e., containing filter 10) into satellite container 52 along conduit 74, connector 84, conduit 75, and conduit 76. In some embodiments, satellite container 51 contains a red blood cell additive solution, that is subsequently passed from container 51 into satellite bag 53, containing red blood cells therein. If desired, the red blood cells, typically combined with additive solution, can be stored until needed.

Figure 5A and 5B illustrate other embodiments of a biological fluid processing system including filter device, and at least one first satellite container suitable for receiving the filtered fluid passing through the filter device. Accordingly, Figure 5A

shows filter device 100 and a downstream first satellite container 51, in fluid communication via conduits 90 and 91, and Figure 5B shows filter device 100 and two first satellite containers 51a and 51b, in fluid communication via conduits 92-95 and connector 81. The embodiments illustrated in Figures 5A and 5B are especially suitable for attachment (for example, via sterile docking) to other biological fluid processing systems, e.g., including collection and/or satellite bags (including wet bag systems) and/or apheresis systems.

The embodiment illustrated in Figure 5B (i.e., including two first satellite containers 51a and 51b) is especially useful for use with some apheresis systems, for example, those systems that provide for collecting two units of biological fluid.

In exemplary variations of the system illustrated in Figure 4, the system does not include leukocyte filter device 200 and third satellite container 53, or the system includes third satellite container 53 but does not include leukocyte filter device 200. However, if desired, the additional system component(s) can be added, e.g., via sterile-docking.

EXAMPLE 1

A system is arranged as generally shown in Figure 4.

A filter comprising 6 first filter elements and 5 second filter elements is placed in a housing to provide a filter device. The first and second filter elements alternate, to provide a filter having 11 layers. The first filter element is utilized for layers 1, 3, 5, 7, 9, and 11, and the second filter element is utilized for layers 2, 4, 6, 8, and 10. Each filter element is a planar circular disc having a diameter of about 47 mm.

The 11 layers are uncompressed, and the resultant filter has an average voids volume of about 91%.

The first filter element and second filter element each comprise melt-blown PBT fibers.

The first filter element, comprising a leukocyte depletion medium, wherein the element does not substantially activate C3a, is surface modified in accordance with U.S. Patent No. 4,880,548, and has a CWST of 95 dynes/cm, and a negative zeta potential at a

pH of about 7.1.

The second filter element, comprising a leukocyte depletion medium wherein the element substantially removes platelets, is surface modified with gas plasma in accordance with U.S. Patent No. 5,258,127, and has a CWST of 65 dynes/cm, and a negative zeta potential at a pH of about 7.1.

Units of whole blood are collected in collection bags containing citrate-phosphate-dextrose preservative, i.e., CPD or CP2D. Each collection bag has a top port and a bottom port. The collection bag containing the blood therein is centrifuged to provide three fractions of blood components, i.e., a sediment layer comprising concentrated red blood cells, an intermediate layer comprising the "buffy coat," and a supernatant layer comprising platelet-poor-plasma (PPP).

The collection bag is placed in a plasma expressor and the PPP is expressed from the top port of the collection bag, through the filter device, and into an empty satellite bag. Red blood cells are not visible in the fluid passing into the satellite bag. Flow is stopped when about 50 ml of plasma remain in the bag, e.g., before the buffy coat is passed into the satellite bag. The filtration time is about 6 minutes.

Analysis of the unit of filtered PPP shows less than 57 white cells in the total volume of PPP (corresponding to less than about 200 white blood cells per liter). There are less than about 650 platelets/μL present.

The C3a concentration in the filtered PPP is below the detection limit of the assay, that is 137.5 ng/ml.

This example shows that filter devices according to the invention can provide substantially leukocyte-free and substantially C3a-free plasma.

EXAMPLE 2

Filter devices are provided as described in Example 1. Units of apheresed plasma are obtained, and filtered within approximately 1 hour of collection. The average volume of the units is about 650 ml. The filter device is interposed between the bag containing the unit of apheresed plasma, and an empty satellite bag, and the plasma-containing bag and the filter device are arranged to allow gravity filtration, with

a head height of 60 inches.

The average filtration time is 16 minutes. Analysis of the filtered plasma shows the average leukocyte residual is 1.8×10^4 white blood cells per unit, and the average platelet residual is 1.8×10^9 platelets per unit. The C3a concentration in the filtered plasma is below the detection limit of the assay.

This example shows that filter devices according to the invention can filter a unit of apheresed plasma in a suitable amount of time while providing substantially leukocyte-free and substantially C3a-free plasma.

EXAMPLE 3

A filter is configured having 11 layers of PBT as described in Example 1. The first filter element, having 6 layers, is surfaced modified by treatment with hydroxyethyl methacrylate (HEMA) and methacrylic acid (MAA), as generally disclosed in U.S. Patent No. 4,880,548.

The second filter element, having 5 layers, is surface modified by treatment with a mixture of argon and ammonia gas plasma as generally disclosed in U.S. Patent No. 5,258,127.

Samples of unmodified PBT, and samples from the first and second elements are analyzed by X-ray photoelectron spectroscopy (XPS), using a Physical Electronics 5700LSci ESCA spectrometer. The X-ray source is monochromatic aluminum, the source power is 350 watts, the exit angle (the angle between the surface plane and the electron analyzer lens) is 50°, and the charge correction is C-(C,H) in C 1s spectra at 284.6 eV.

The concentration of elements detected (in Atom %) are shown below in Table I, and the summary of carbon functional groups (in Atom % Carbon) are shown below in Table II.

Concentration of Elements Detected (in Atom %)

sample	O	N	C	Si
unmodified	23.7	0.2	76.1	0.1
HEMA/MAA	28.0	0.5	71.4	0.0
gas plasma	18.8	3.1	78.0	0.1

Table I

Summary of Carbon Functional Groups (in Atom % Carbon)

	C-(C,H)		C-(O,N)[@]		C=O[#]		(O,N)-C=O[$]	
sample	B.E.[+]	atom% C	B.E.[+]	atom% C	B.E.[+]	atom% C	B.E.[+]	atom% C
unmod[*]	284.6	54	286.0	11	-	-	288.5	10
HEMA /MAA	284.6 [&]	49	286.1	12	287.2	3	288.5	5
gas plasma	284.6	58	286.1	13	-	-	288.4	7

+: binding energy
@: alcohol, ether, amine
#: ketone, aldehyde
$: ester, carboxylic acid, amide
*: also contained about 1 atomic% aromatic carbon (e.g., phenyl, benzyl)
&: the C-(C,H) peak is a combination of two peaks

Table II

The data shows that an embodiment of a filter according to the invention has a first element having a surface that is hydroxylated relative to its bulk, and a second element having a surface having a nitrogen-to-oxygen ratio of 0.017.

EXAMPLE 4

A filter is prepared as described in Figure 3, i.e., the first and second filter elements (comprising melt-blown PBT) alternate, to provide a filter having 11 layers. The first filter element is utilized for layers 1, 3, 5, 7, 9, and 11, and the second filter element is utilized for layers 2, 4, 6, 8, and 10. Each filter element is a planar circular disc having a diameter of about 47 mm. The first filter element has a CWST of 95 dynes/cm and a zeta potential of -8 mv at a pH of 7.0. The second filter element has a CWST of 65 dynes/cm, and a zeta potential of -10 mv at a pH of 7.0.

The average fiber diameter of the fibers in each element is about 2.7 micrometers.

The 11 layers are calendered together utilizing heat compression on a continuous belt. The resultant laminate filter has an average voids volume of about 79%, and a pore diameter of about 2 micrometers.

Units of whole blood (about 450 mL) are collected, and centrifuged, as generally described in Example 1. Using head heights of 100 cm or 150 cm (equal numbers of units of platelet-poor-plasma are filtered at the two head heights) , platelet-poor-plasma is passed through the filter devices, and flow is stopped before buffy coat passes from the collection container. The filtration times average less than 5 minutes.

Analysis of the filtered fluid shows, on the average, less than 150 white blood cells per unit of PPP (corresponding to less than 500 white blood cells per liter). On the average, there are less than 5×10^9 platelets per unit. The C3a concentration of the filtered plasma is below the detection limit of the assay.

This example shows filter devices according to an embodiment of the invention can be used to provide substantially leukocyte-, platelet- and C3a-free plasma via gravity filtration.

All of the references cited herein, including publications, patents, and patent applications, are hereby incorporated in their entireties by reference.

While the invention has been described in some detail by way of illustration and example, it should be understood that the invention is susceptible to various

modifications and alternative forms, and is not restricted to the specific embodiments set forth. It should be understood that these specific embodiments are not intended to limit the invention but, on the contrary, the intention is to cover all modifications, equivalents, and alternatives falling within the spirit and scope of the invention.

WHAT IS CLAIMED IS:

1. A filter for processing a biological fluid comprising:

at least two filter elements, wherein the surface of one filter element has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00, and the surface of the other filter element is hydroxylated relative to the bulk of the element.

2. The filter of claim 1, further comprising at least one additional filter element, wherein the surface of the additional element has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00.

3. The filter of claim 1, further comprising at least one additional filter element, wherein the surface of the additional element is hydroxylated relative to the bulk of the element.

4. The filter of claim 1, further comprising at least two additional filter elements, wherein the surface of the first additional element has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00, and the surface of the second additional element is hydroxylated relative to the bulk of the element.

5. The filter of claim 2, wherein the element having the hydroxylated surface is interposed between the two elements having surfaces including the nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00.

6. The filter of claim 3, wherein the element having a surface including the nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00 is interposed between the two elements having hydroxylated surfaces.

7. A filter for processing a biological fluid comprising:

at least one filter element comprising a porous medium, wherein at least a portion of the surface of the element is aminated and hydroxylated relative to its bulk.

8. A filter for processing a biological fluid comprising:

at least one filter element comprising a porous medium, wherein a portion of the surface of the element is aminated relative to the bulk of the element, and another portion of the surface of the element is hydroxylated relative to the bulk of the element.

9. A filter device for processing a biological fluid comprising:

a housing having an inlet and an outlet and defining a fluid flow path between the inlet and the outlet;

a filter disposed in the housing across the fluid flow path, the filter comprising:

at least one filter element comprising a porous fibrous leukocyte depletion medium having a first predetermined CWST; and

at least one filter element comprising a porous fibrous leukocyte and platelet depletion medium having a second predetermined CWST;

wherein the filter is arranged to allow plasma to pass therethrough and substantially prevent the passage of leukocytes and platelets therethrough.

10. A filter device for processing a biological fluid comprising:

a housing having an inlet and an outlet and defining a fluid flow path between the inlet and the outlet;

a filter disposed in the housing across the fluid flow path, the filter comprising:

at least one filter element comprising a porous fibrous leukocyte depletion medium having a first predetermined CWST; and

at least one filter element comprising a porous fibrous leukocyte depletion medium having a second predetermined CWST;

wherein the filter is arranged to allow plasma to pass therethrough and substantially prevent the passage of leukocytes therethrough, without substantially activating C3a in the biological fluid.

11. A filter device for processing a biological fluid comprising:

a housing having an inlet and an outlet and defining a fluid flow path between the

inlet and the outlet;

a filter disposed in the housing across the fluid flow path, the filter comprising:

at least one filter element comprising a porous fibrous leukocyte depletion medium having a first predetermined CWST; and

at least one filter element comprising a porous fibrous leukocyte and platelet depletion medium having a second predetermined CWST;

wherein the filter is arranged to allow plasma to pass therethrough and substantially prevent the passage of platelets, leukocytes and C3a therethrough.

12. The filter of claim 1, or the filter device of any of claims 9-11, wherein the first filter element has a CWST that is different than the CWST of the second filter element.

13. The filter or filter device of any preceding claim wherein the filter is arranged to allow plasma to pass therethrough and substantially prevent the passage of platelets, leukocytes and red blood cells therethrough.

14. The filter or filter device of any preceding claim wherein the filter includes at least one additional filter element, the filter element comprising:

a porous fibrous leukocyte depletion medium having the first predetermined CWST; and/or

a porous fibrous leukocyte and platelet depletion medium having the second predetermined CWST.

15. The device according to claim 14, wherein the filter includes a porous fibrous leukocyte depletion medium having the first predetermined CWST interposed between a first porous fibrous leukocyte and platelet depletion medium having the second predetermined CWST, and a second porous fibrous leukocyte and platelet depletion medium having the second predetermined CWST.

16. The device according to claim 14, wherein the filter includes a porous fibrous

leukocyte and platelet depletion medium having the second predetermined CWST interposed between a first porous fibrous leukocyte depletion medium having the first predetermined CWST, and a second porous fibrous leukocyte depletion medium having the first predetermined CWST.

17. The device of claim 12, wherein the filter includes at least one additional filter element, the filter element comprising:

a porous fibrous leukocyte depletion medium having the first predetermined CWST; and/or

a porous fibrous leukocyte and platelet depletion medium having the second predetermined CWST.

18. The filter or filter device of any of the preceding claims, wherein at least one filter element comprises at least two layers.

19. The device of any of the preceding claims, wherein at least one filter element has a CWST of at least about 90 dynes/cm.

20. The filter or filter device of any of the preceding claims, wherein the filter includes at least two filter elements, wherein the surface of one filter element has a nitrogen-to-oxygen ratio in the range of from at least 0.01 to less than about 1.00, and the surface of the other filter element is hydroxylated relative to its bulk.

21. The filter or filter device of claim 20, wherein the surface of one filter element has a nitrogen-to-oxygen ratio in the range of from at least about 0.02 to less than about 1.00, and the surface of the other filter element is hydroxylated relative to its bulk.

22. The filter or filter device of any of the preceding claims, wherein the filter is arranged to provide leukocyte-depleted plasma having about 1×10^3 leukocytes or less therein.

23. The filter or filter device of any of the preceding claims wherein the filter is arranged to provide platelet-depleted plasma having about 1 x 10^9 platelets or less therein.

24. The filter or filter device of any of the preceding claims, wherein the filter substantially removes C3a from the biological fluid passing therethrough.

25. The filter or filter device of any of the preceding claims, wherein C3a is not substantially activated by the filter as the biological fluid passes through the filter.

26. A method for processing a biological fluid comprising:

passing a biological fluid through the filter or filter device of any proceeding claim, and

obtaining the filtered fluid.

27. A method for processing a biological fluid comprising:

passing a leukocyte-containing plasma-rich biological fluid through a filter comprising at least one filter element, wherein at least a portion of the surface of the element is aminated relative to its bulk, and another portion of the surface of the element is hydroxylated relative to the element's bulk; and obtaining a filtered plasma-rich biological fluid substantially free of leukocytes and platelets.

28. The method of claim 27, wherein the filter comprises at least two filter elements, and at least a portion of the surface of one element is aminated relative to its bulk, and at least a portion of the surface of the other element is hydroxylated relative to its bulk.

29. A method for processing a biological fluid comprising:

passing a leukocyte-containing plasma-rich biological fluid through a filter device comprising a filter including:

a first element comprising a porous fibrous leukocyte depletion medium having a first predetermined CWST; and

a second element comprising a porous fibrous leukocyte depletion medium having a second predetermined CWST; and

collecting a filtered plasma-rich biological fluid substantially free of leukocytes and platelets.

30. A method for processing a biological fluid comprising:

passing a leukocyte-containing plasma-rich biological fluid through a filter device comprising a filter including a fibrous leukocyte depletion medium and a fibrous leukocyte and platelet depletion medium; and

collecting a filtered plasma-rich biological fluid substantially free of leukocytes, and C3a.

31. A method for processing a biological fluid comprising:

passing a leukocyte-containing plasma-rich biological fluid through a filter comprising at least one filter element, wherein at least a portion of the surface of the element is aminated and hydroxylated relative to its bulk; and obtaining a filtered plasma-rich biological fluid substantially free of leukocytes and platelets.

32. A method for processing a biological fluid comprising:

passing a platelet-poor plasma-rich biological fluid through a filter device comprising a filter including a fibrous leukocyte depletion medium and a fibrous leukocyte and platelet depletion medium; and

collecting a filtered plasma-rich biological fluid substantially free of platelets and leukocytes.

33. The method of any of the preceding claims wherein the filtered plasma-rich fluid is substantially free of C3a.

34. The method of any of the preceding claims wherein the leukocyte-containing plasma-rich biological fluid comprises a platelet-poor biological fluid.

35. The method of any one of claims 24-34, including collecting plasma-rich fluid in a downstream container without substantially activating C3a in the plasma-rich fluid.

36. A system comprising the filter or filter device of any one of claims 1-23, and further comprising a container downstream of the filter or filter device, and in fluid communication therewith.

37. The system of claim 36, comprising a closed system.

FIG. 1

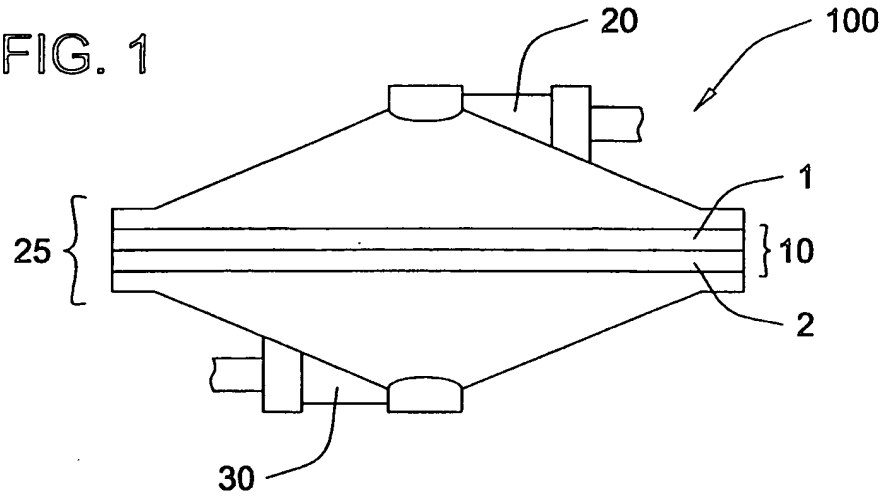


FIG. 2

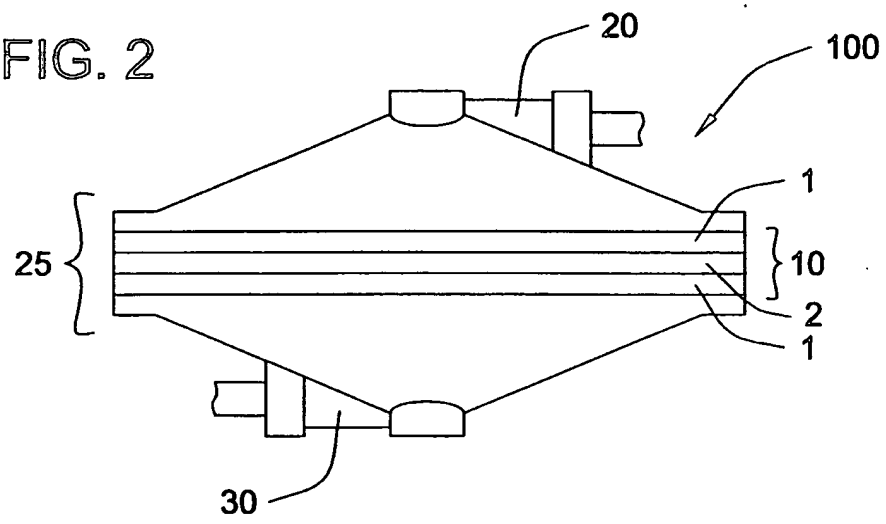
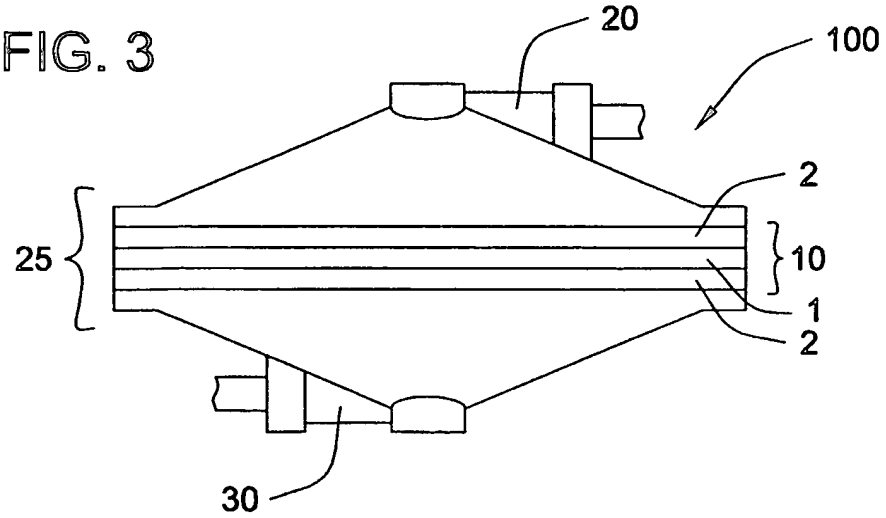


FIG. 3



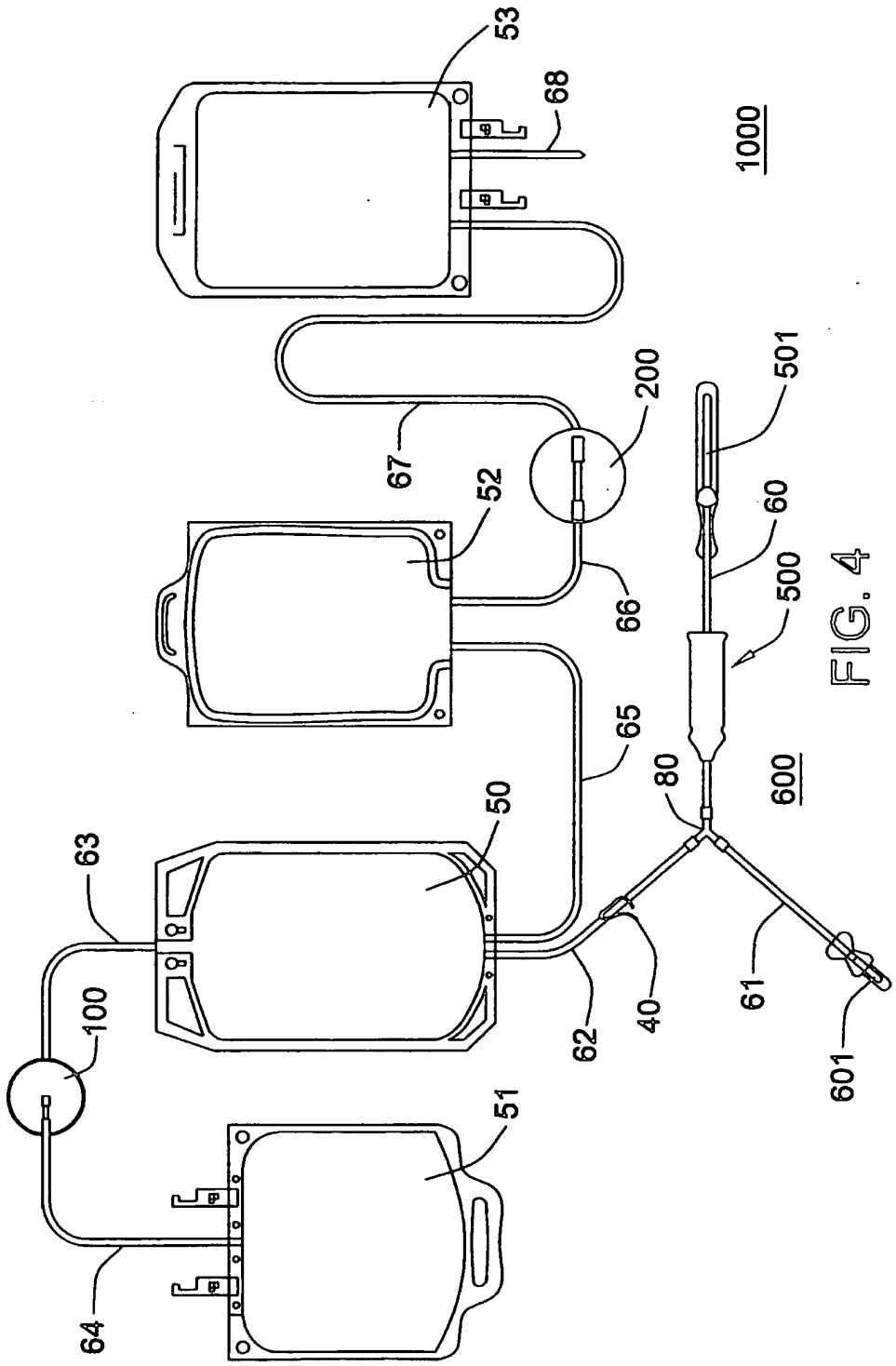

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FIG. 4

FIG. 5B

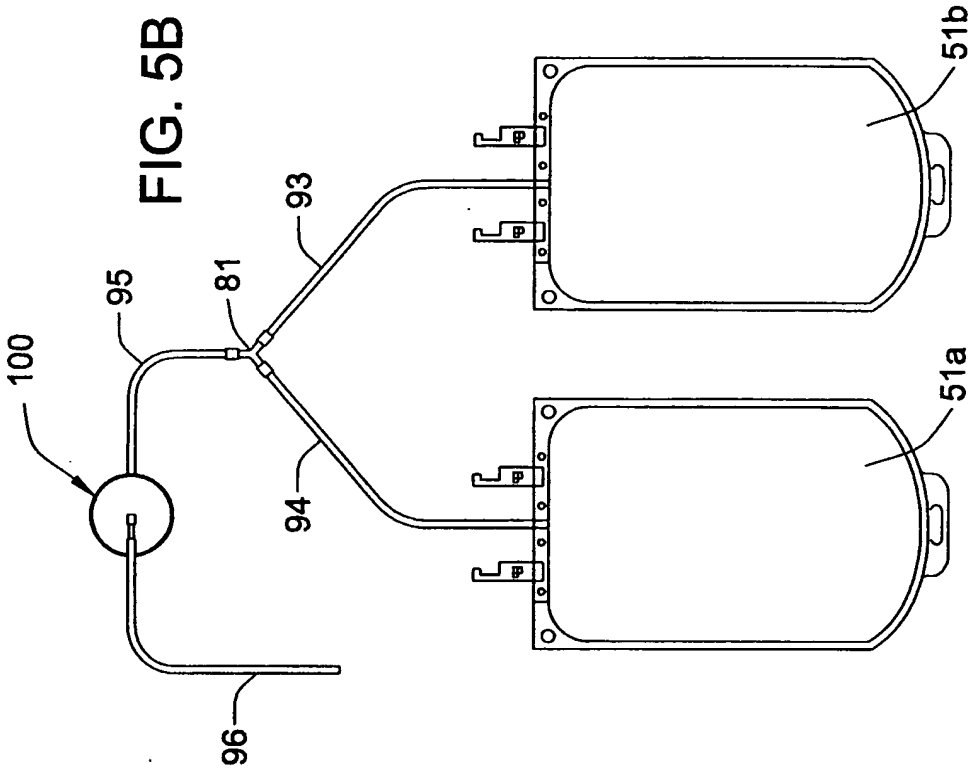
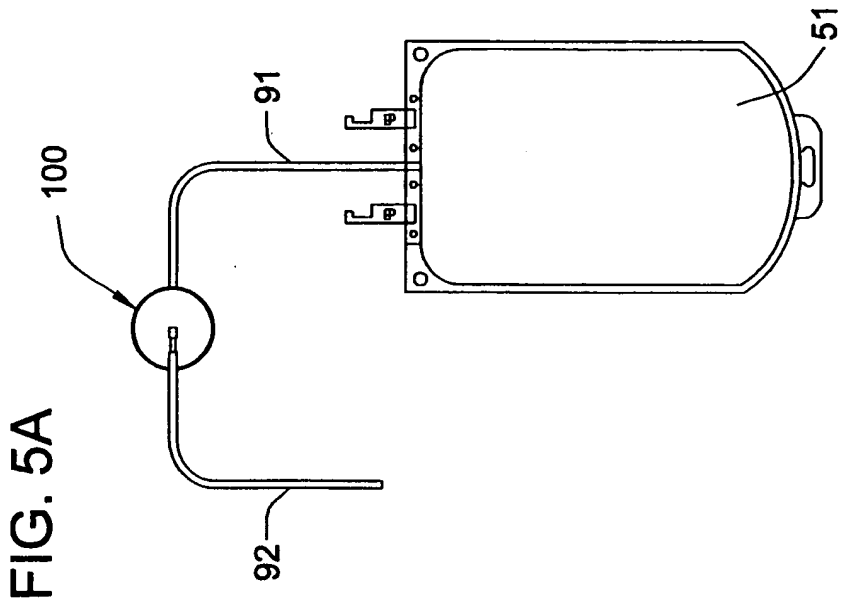


FIG. 5A



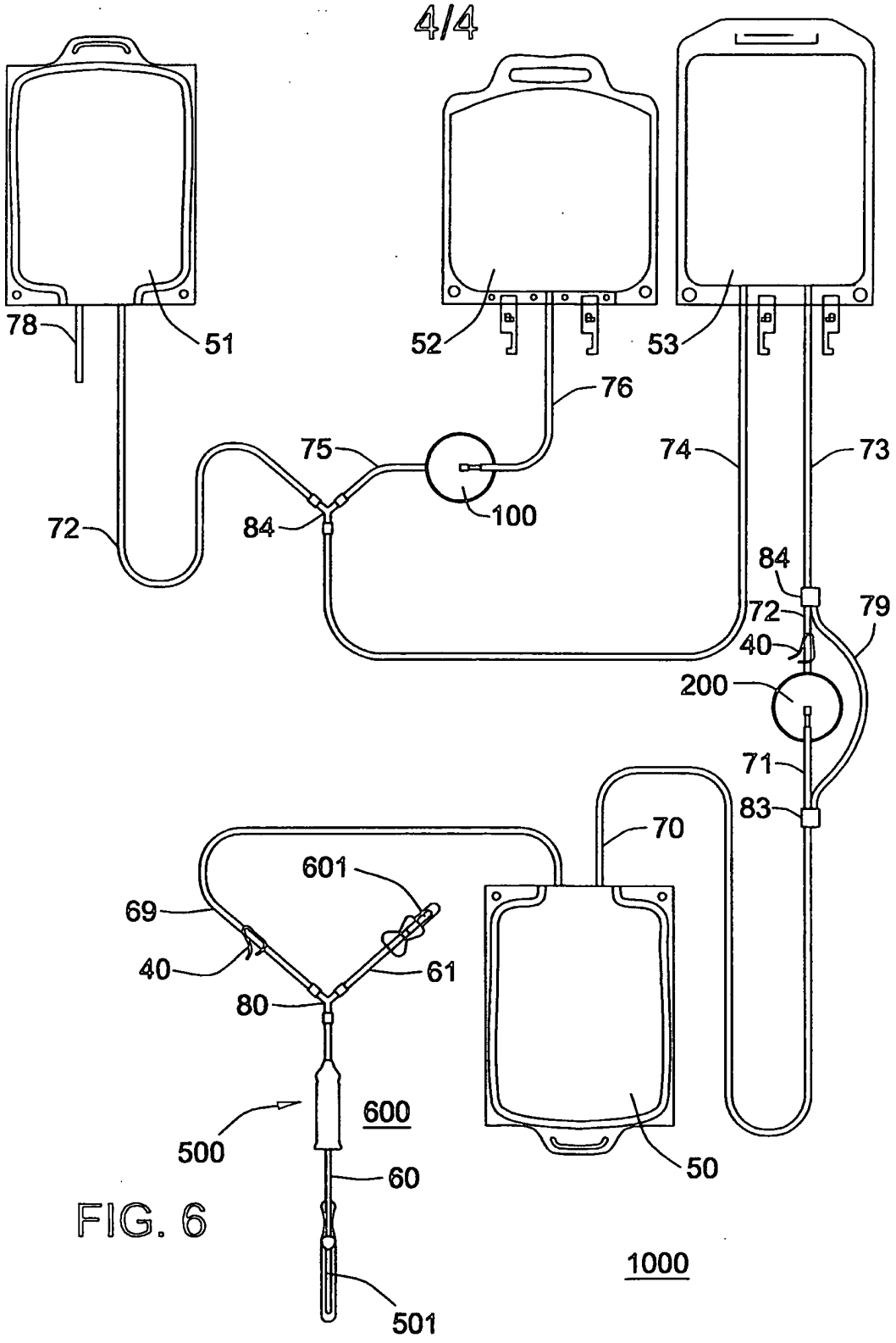

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FIG. 6